Journal of Agricultural Chemistry and Biotechnology

Journal homepage: <u>www.jacb.mans.edu.eg</u> Available online at: <u>www.jacb.journals.ekb.eg</u>

Antioxidant Activity of Some Natural Compounds in Alleviating the Hepatotoxicity Effects Induced by Emamectin Benzoate in Male Mice

Sherifa F. M. Dawoud ¹; T. M. M. Al-Akra² and Amina M.G. Zedan^{1*}

¹Biological and Environmental Sciences Dept. Faculty of Home Economic, Al-Azhar University ²Agric. Zoology and nematology Dept. Faculty of Agric., Al-Azhar University



ABSTRACT



Emamectin benzoate (EMB) is a biopesticide that is used in agriculture as an insecticide. Due to its wide range of use, it is easier to reach ecologically and affects human health. This study aims to evaluate the protective effect of natural compounds against EMB-induced hepatotoxicity. It is the first study quantifying the hepatoprotective effect of these extracts against EMB effects. Biochemical analysis includes MDA, CAT, SOD, ALT and AST analysis. However, genetic studies include the gene expression of Mgst1, CYP2E1, caspase3, IL-16 and DNA fragmentation, as well as, histopathological investigations were performed. Male mice were distributed into five groups: G1: the negative control, G2: EMB group (5mg/kg diet), G3: EMB+Boswellia serrata (90 mg/kg diet), G4: EMB+Cinnamonum zeylanicum (600mg/kg diet), G5: EMB+ powder of the snail (600mg/kg diet), and the experiment continues for eight weeks. The results appeared that EMB induced oxidative stress in the liver by increase the activity of MDA. The biomarker of liver injury ALT and AST were elevated with antioxidant enzymes inhibition. EMB produced several histopathological changes in the liver. Relative expressions of Mgst1, CYP2E, caspase-3 and IL-1ß genes elevated in the liver. The increase in DNA damage was noticed as recorded by an increase in tail length, tail DNA% and tail moment. Co-treatment with natural compounds reduced the toxicity of EMB. They reduce the abnormal biochemical, histopathological, gene expression and DNA damage by increasing antioxidant capacity. Therefore, the natural compounds used in this study act as potent hepatoprotective agents against EMB induced hepatotoxicity in mice.

Keywords: Emamectin benzoate; hepatotoxicity; gene expression; DNA fragmentation; antioxidants

INTRODUCTION

Pesticides used in public health or in agriculture sector have various actions on metabolic mechanisms. It affects on non-target animal and human health (El-Bialy et al., 2020). One of macrocyclic lactone is emamectin benzoate (EMB). It is developed as a pesticide formed by avermectins fermentation. The chemical structure contains 10% avermectin B1b and 90% avermectin B1a (Hayes and Laws 2013). In the human central nervous system, γ aminobutyric acid-reactive neurons are limited so EMB was considered safe to human (Roberts et al., 1984), but the EMB-lipophilicity makes it penetrates the membranes of the cell. Consequently, it creates toxicity in animals and humans (Wolterink et al., 2011). Although the data about the effect of EMB on antioxidant status is insufficient. Many studies confirmed that avermectins insecticides created oxidative stress in intoxicated animals. Similarly, it created the kidney and liver oxidative stress in offspring and mothers after lactational exposure (Mossa et al., 2017). One of the mainly valuable medicinal plants is Boswellia serrata (BS) (Mark, 2018). The treatment with BS significantly leads to a decrease in liver fibrosis. BS has the ability for recovering endogenous antioxidant mechanisms. Subsequently, it scavenges free radicals and allows hepatocyte regeneration (Eltahir et al., 2019). Cinnamomum zeylanicum (CZ) prevents and treats many diseases (Hussain et al., 2019). Cinnamon is utilized extensively as an herbal drug (Morgana et al., 2014). It has been applied to the remedy of

gastric diseases and inflammatory disorders (Shen *et al.*, 2012). Phytochemical study of cinnamon has detected a lot of coumarins, flavonoids, alkaloids, glycosides, steroids, anthraquinone, terpenoids and tannins (Shihabudeen *et al.*, 2011). Cinnamon's hepatoprotective impact has been reported by Eidi *et al.* (2012). Garden snails are one of invertebrates. To date, it has given a wide variety of natural products. It is including terpenes, aliphatic hydrocarbons, alkaloids, steroids, amino acids, peptides, and carbohydrates (Leal *et al.*, 2012). The natural products of snails have a wide array of remedial properties including; antimicrobial, anticoagulant, immune-modulating, wound healing, anticancer, antioxidant, antihypertensive and anti-inflammatory (Perdicalis *et al.*, 2013).

Consequently, the aim of this study was to evaluate the protective role of some natural compounds against oxidative stress and hepatotoxicity induced by EMB in mice.

MATERIALS AND METHODS

Emamectin benzoate (EMB) have a trade name called Speedo (5.7% WG) was purchased from EL-Shoura chemicals Co., was used in this study as insecticide.

Plant material and extraction

For preparing methanol extract of *Boswellia serrata* and *Cinnamomum zeylanicum*, the methods of Yazdanpanahi, *et al.* (2014) and Ojarudi, *et al.* (2020) were used respectively with some modifications belong to the concentrations. The cinnamon bark and gum resin of *Boswellia serrata* were obtained from the local market.

They were crushed into small pieces. Two hundred grams were placed in a flask and macerated with 600 mL methanol in a water bath with a shaker for 2 hours at 50 °C. Then, the mixture was filtered with Whatman No. 1 filter paper more than once and dried at room temperature. The crude extract was stored in a well-closed container, protected from light, and kept at 4° C.

Preparation snail's powder

The garden snails (*Eobania vermiculata*) were dried in the oven at 50-60 0 C for two hours. Then ground in a blender to a very fine powder.

Chemical Composition

The chemical composition of natural extracts and snail's powder was estimated. Bioactive compounds are measured by GC/MS System (Thermo Scientific TRACE 1310 Gas Chromatograph, USA) attached with an ISQ LT single quadrupole mass spectrometer according to Hadi and Hameed (2017) at The Institute of Marine Biology research, The National Research Center

Animals and groups

In this experiment, 45 male albino mice with an approximate weight of 19 gm were used. The animals were maintained in the standard condition (temperature $22 \pm 2^{\circ}$ C, with a light/dark cycle of 12 hr) in stainless steel cages in an artificially illuminated. After compatibility, the animals were divided randomly into five groups, each containing nine animals for each treatment. They were treated as follows; G1: mice are given a diet without any additives, G2: mice are treated with EMB at a dose of 5 mg/kg (National Registration Authority for Agricultural and Veterinary Chemicals, 2011) diet (equivalent to 1/10 LD50), G3: mice are treated Boswellia (90 mg/kg diet) (Yassin et al., 2013) with EMB, G4: mice are treated Cinnamomum (600 mg/kg diet) (Prasanna and Anand 2019) with EMB, G5: mice are treated powder of the snail (PS) (600 mg/kg diet) with EMB (this is the first study on PS). After eight weeks mice have fasted overnight. They were sacrificed after anesthetized using diethyl ether and their organs (liver, kidney, spleen, heart, and testis) were removed and immediately weighed after sacrifice. The liver was quickly removed and stored in liquid nitrogen until transferred to -80 and then used for genetic studies. Another portion of liver stored in formalin 10% for histological studies.

Biochemical studies

Blood samples were taken in a glass tube without EDTA and left for two hours for coagulating at room temperature. It was centrifuged at 4000 rpm for 20 min to obtain sera samples. Serum samples were kept at -20° C until used for biochemical assays. According to the technique of Reitman and Frankle (1957) AST and ALT activity were measured. Malondialdehyde (MDA) was investigated according to the technique of Ohkawa *et al.* (1979). Catalase (CAT) was estimated according to Aebi (1984). Superoxide dismutase (SOD) was estimated according to Nishikimi *et al.* (1972).

Histopathological examination

Small parts of the liver were fixed in formalin solution (10%). They dehydrated in ethanol from 70% to 100%. Then, they cleared in xylene and transfer to paraffin. The sections of liver stained with Eosin and Hematoxylin dyes (Suvarna *et al.*, 2013).

Molecular analysis

The RNeasy kit (Qiagen) was used for isolated total RNA from the liver as described by Abd-Allah *et al.* (2015). The purity and integrity of RNA were assessed by Nanodrop, and 1% agarose gel electrophoresis, respectively. The Quantiscript reverse transcriptase was used in RNA reverse transcription to cDNA. Real-time PCR reaction contains cDNA as a template in the presence of QuantiTect SYBR Green qPCR Master Mix and gene-specific primers, designed by the Primer 3 web-based tool based on the published mouse sequence (Table 1), along with Step One Plus real-time PCR system (Applied Biosystem, USA) and reaction cycles as described by Khamis *et al.* (2018). The critical threshold (Ct) quantities for the target genes were normalized with quantities of the Ct of the internal control (β -actin).

Table1. Primers used for	or real-time PCR.
--------------------------	-------------------

I upiciti I i i i i i i i i i i i i i i i i i	Tubler: I Timer's used for rear time r erk							
Gene	Forward primer (/5 /3)	Reverse primer (/5 /3)						
Mgst1	TTTTGCCAACCCGGAAGACT	GAGGCCGATACCGAGAAAGG						
Cyp2E1	CTCCTCGTCATATCCATCTG	GCAGCCAATCAGAAATGTGG						
Caspase 3	GGTATTGAGACAGACAGTGG	CATGGGATCTGTTTCTTTGC						
IL1b	CACCTCTCAAGCAGAGCACAG	GGGTTCCATGGTGAAGTCAAC						
β -actin	AAGTCCCTCACCCTCCCAAAAG	AAGCAATGCTGTCACCTTCCC						

Comet assay

Comet assay was performed according to Eldamaty *et al.* (2021). In brief, a weight of 1 gram of crushed samples was transferred to 1 ml ice-cold PBS. This suspension was stirred for 5 min and filtered. 100 μ 10 f cell suspension was integrated with 600 μ 1 of agarose (low-melting, 0.8% in PBS). This mixture (100 μ 1) was spread on slides. The slides were immersed in lysis buffer for 15 min. Then they were placed in the electrophoresis chamber containing the same lysis buffer without SDS. The conditions of electrophoresis were 100 mA and 2 V/cm for 2 min. Staining with ethidium bromide. The DNA fragment migration patterns were evaluated with a fluorescence microscope at a magnification of 40x and with excitation filter 420-490nm. Komet 5 image analysis software

developed by Kinetic Imaging, Ltd. (Liverpoo1, UK) attached to a CCD camera. It was used to assess the qualitative and quantitative extent for DNA damage by calculates the DNA-migration length and the percentage of migrated DNA.

Ethics approval

This experiment was carried out under Egyptian ethical codes for studies on experimental animals and approved by the Ethics Committee of Al-Azhar University. The experimental protocol was approved by the Biological and Environmental Sciences Department, Faculty of Home Economics, Al-Azhar University, Egypt.

Statistical analysis

All the data were expressed as means ±SD. The statistical significance was evaluated by one-way ANOVA

(analysis of variance) using SPSS version 20 software, and the individual comparisons were obtained by Duncan's multiple range test (DMRT). Values were considered statistically significant when p<0.05 (Bryman and Cramer, 2011).

RESULTS AND DISCUSSION

GC-MS analysis

The chemical compositions of methanol extract for *Boswellia serrata* are shown in Table 2. Fourteen compounds were identified. The major comprised organic compounds were 1,6,10-Dodecatrien-3-ol,3,7,11 trimethyl-, (E)- (16.19%) at RT 17.9 that acts as antidiabetic, hepatoprotective and anti- inflammatory activities. Also it contains 1-Heptatriacotanol that has antioxidant, anticancer, anti inflammatory and sexhormone activity (Shareef *et al.*, 2016) and this compound present in BS, CZ and PS (Kalairasan *et al.*, 2011). The major components of the essential oils of the Boswellia found in this study is Isopropyl-1,5,9-trimethyl-15 oxabicyclo[10.2.1]pentadeca-

5,9 dien-2-ol (Isoincensole) with concentration 58.30% (Awoke and Joshi, 2021).

Table (3) illustrated that there are twenty one of bioactive compounds in Cinnamon, the major are 1-[1-(2,2 Dichlorovinylimino)-2,2-di methylpropyl]-3-(p tolyl)thiourea (7.30%), 1-Heptatriacotanol (8.51%) and 24-Norursa-3,12-diene (8.43%). The bioactive compounds presented in Cinnamomum that played a major role in reducing the side effect of EMB as Oleic acid that found in CZ and PS and have anti-inflammatory, anti-androgenic, preservative hypocholesterolemic anti-cancer, and characters (Sreekumar et al., 2014). PS also contain bioactive compounds that have antiinflammatory, hypocholesterolemic, cancer preventive, and hepatoprotective effects such as 12,15-Octadecadienoic acid, methyl ester, (Z,Z,Z)- (Rehana and Nagarajan 2013). The bioactive compounds of PS are showing in Table (4). There are twenty bioactive compounds, the major compounds are 2-(2 butoxyethoxy) ethyl acetate (11.49%) and benzene, 1,2,4-trimethyl- (21.39%).

Table 2. Bioactive com	pounds identified ii	n methanolic extract of	Boswellia serrata

n	RT	Compound Name	Area %	Peak Area	Formula	MW
1	7.48	Cyclopropane, pentyl	2.52	1711415172.16	C8H16	112
2	10.20	1-HEXANOL, 2-ETHYL-, ACETATE	4.55	3091011881.26	C10H20O2	172
3	17.90	1,6,10-Dodecatrien-3-ol,3,7,11 trimethyl-, (E)-	16.19	10998543245.16	C15H26O	222
4	18.27	Dodecanoic Acid, Ethyl Ester	0.98	662240817.76	C14H28O2	228
5	24.32	1,3,6,10 Cyclotetradecatetraene, 3,7,11-trimethyl-14-(1 methylethyl)-, [S-(E,Z,E,E)]-	1.01	686677160.95	C20H32	272
6	24.85	(R,1E,5E,9E)-1,5,9-Trimethyl-12-(prop-1-en-2-yl)cyclotetradeca-1,5,9- triene	2.40	1630904366.91	C20H32	272
7	25.31	Hexadecanoic acid, ethyl ester	0.81	550653815.15	C18H36O2	284
8	25.75	(S,E)-8,12,15,15-Tetramethyl-met hylenebicyclo[9.3.1]pentadeca-7,11-diene	1.27	859650907.34	C20H32	272
9	26.45	Thunbergol	6.72	4564371557.29	C20H34O	290
10	27.00	1-Heptatriacotanol	1.25	845938343.56	C37H76O	536
11	27.91	(3E,7E,11E)-1-Isopropyl-4,8,12trime thylcyclotetradeca-3,7,11-trienol	1.16	789102248.63	C20H34O	290
12	28.17	Isopropyl-1,5,9-trimethyl-15 oxabicyclo[10.2.1]pentadeca-5,9 dien-2-ol	58.30	39595790847.85	C20H34O2	306
13	30.12	Nerolidol-Epoxyacetate	1.90	1287616100.01	C17H28O4	296
14	37.70	15,17,19,21 Hexatriacontatetrayne	0.94	641545071.66	C36H58	490

Table 3. Cinnamon zeylanicum-bioactive compounds identified in methanolic extract

n	RT	Compound Name	Area %	Peak Area	Formula	MW
1	4.14	1-[1-(2,2 Dichlorovinylimino)-2,2-di methylpropyl]-3-(p tolyl) thiourea	7.30	60352088.42	C15H19Cl2N3S	343
2	4.23	12,15-Octadecadiynoic Acid, Methyl Ester	5.95	49209036.43	C19H30O2	290
3	4.78	Acetic acid, Octyl Ester	2.64	21792402.94	C10H20O2	172
4	4.87	1-Deoxy-d-arabitol	5.27	43599137.66	C5H12O4	136
5	5.38	Benzene, 1-Ethyl-3-Methyl-	4.24	35021407.31	C9H12	120
6	5.94	Benzene, 1,2,3-Trimethyl-	5.19	42936038.66	C9H12	120
7	6.49	Butanoic acid,2-amino-4 (methylsulfinyl)-, (ñ)-	3.10	25661984.56	C5H11NO3S	165
8	7.73	2-Myristynoyl pantetheine	3.79	31359300.08	C25H44N2O5S	484
9	9.11	2-t-Butyl-5-propyl [1,3]dioxolan-4-one	4.28	35417344.02	C10H18O3	186
10	11.84	Isobornyl thiocyanoacetate	2.79	23044475.82	C13H19NO2S	253
11	24.32	9-Octadecenoic acid (Z)-	4.67	38621464.83	C18H34O2	282
12	27.04	9-Octadecenoic acid (Z)-	6.98	57753324.42	C18H34O2	282
13	28.12	Tetraneurin - A -DIOL	5.50	45458012.84	C15H20O5	280
14	28.48	9-Octadecenoic acid (Z)-	3.25	26853996.60	C18H34O2	282
15	28.55	12-Methyl-E,E-2,13-octadecadien-1-ol	1.25	10295788.30	C19H36O	280
16	28.61	1,2,3-propanetriyl ester, (E,E,E)-	4.10	33897683.33	C57H104O6	884
17	30.14	2-Hydroxy-3-[(9E)-9-Octadec Enoyloxy]propyl	3.33	27495030.34	C39H72O5	620
18	30.50	Ethyl iso-allocholate	4.00	33058044.47	C26H44O5	436
19	30.74	1-Heptatriacotanol	8.51	70329474.00	C37H76O	536
20	30.86	24-Norursa-3,12-diene	8.43	69688437.10	C29H46	394
21	31.13	Methyl Commated	5.45	45039447.46	C31H50O4	486

Table 4.	chemical	compounds	identified i	n powder o	of snails
14010 11	citcititeut	compounds	iacitute i	in pomaci u	

n	RT	Compound Name	Area %	Peak Area	Formula	MW
1	4.11	Benzene, 1,2Dimethyl-	4.38	111174139.15	C8H10	106
2	4.28	12,15-Octadecadiynoic acid, methyl ester	1.20	30377255.91	C19H30O2	290
3	4.42	Benzene, 1,2Dimethyl-	5.98	151704650.28	C8H10	106
4	4.73	2-(2 Butoxyethoxy)Ethyl Acetate	11.49	291626987.71	C10H20O4	204
5	4.98	Benzene, 1-Ethyl-3 Methyl-	1.91	48417446.59	C9H12	120
6	5.10	Benzene, 1-ethyl-3-methyl-	2.20	55904135.75	C9H12	120
7	5.95	Benzene, 1,2,4-trimethyl-	21.39	542893530.62	C9H12	120
8	6.46	Benzene, 1,3,5-Trimethyl-	4.47	113545991.43	C9H12	120
9	6.93	Para tolyl Acetaldehyde	1.53	38957845.11	C9H10O	134
10	7.02	4,6-Decadiyne	1.44	36515281.63	C10H14	134
11	7.08	Benzene, 1,2-Diethyl-	2.33	59222824.33	C10H14	134
12	7.72	2-Oxazolamine,	1.22	30939888.78	C17H17N3O3	311
13	8.40	10,13-Octadecadiynoic acid, methyl ester	1.92	48795220.27	C19H30O2	290
14	9.10	7,10 Pentadecadiynoic Acid	1.49	37826282.48	C15H22O2	234
15	24.30	Cyclopropanebutanoic acid,	1.75	44394544.24	C25H42O2	374
16	27.05	9-Octadecenoic Acid (Z)-	5.13	130272311.97	C18H34O2	282
17	28.13	01297107001 Tetraneurin - a -Diol	2.09	52957950.46	C15H20O5	280
18	30.74	24-Norursa-3,12-diene	2.97	75388425.64	C29H46	394
19	30.86	1-Heptatriacotanol	3.26	82663638.05	C37H76O	536
20	31.13	Ethyl iso-allocholate	1.97	50084419.16	C26H44O5	436

EMB effects

The alterations in the body weight (BW) of the mice of all groups and weight of organs are shown in Table. 5. Mice in EMB groups showed a significant decrease (p < 0.05) in the body weight. These results agreed with Khaldoun *et al.* (2015) and El-Sheikha and Gala (2015), they found a decrease in the body weight after treatment with EMB. These decrease due to overstimulation of cholinergic. It causes rise in gastric motility and a reduction in food absorption. These decrease due to reduction in food intake, low palatability of food, or increased lipid and protein degradation due to toxicity related to treatment (Mansour and Mossa, 2010). Treatment with natural compounds effectively alleviate EMB-induced body weight decrease and improves body weight due to they have

and antifibrotic antioxidant, anti-inflammatory characteristics. The treatment with powder of snail alleviates the side effects of EMB that induce body weight decrease. Snails has to date produce a wide diversity of products. These products include alkaloids, terpenes, steroids, aliphatic hydrocarbons (Leal et al., 2012). Significant decrease was obtained in the weight of kidney when the mice treated with EMB if compared to the negative control and other treatments. The negative control recorded the highest weight of kidney. These results indicated that treatment with EMB and other materials did not affect the mean weight of most organs (liver, testes and heart). The weight of spleen was significantly decreased in the treatments if compared with the negative control.

Table 5. Effect of EMB alone or combined with the natural compounds on the body and organs weight of mice (M \pm SD).

Treatment	Body weight	Organs weight					
Treatment	(gm)	Kidney(gm)	Tests(gm)	Liver(gm)	Spleen(gm)	Heart(gm)	
G1	33.78±3.25 ^a	0.29 ± 0.04^{a}	0.13±0.02	1.89±0.36 ^a	0.28±0.02 ^a	0.19±0.03 ^{ab}	
G2	24.08±3.61°	0.21±0.05°	0.12±0.03	1.23±0.49 ^b	0.18 ± 0.04^{b}	0.16±0.03 ^b	
G3	$30.44.\pm 5.17^{ab}$	0.28 ± 0.04^{ab}	0.13±0.01	1.73±0.50 ^{ab}	0.16 ± 0.05^{b}	$0.22 \pm .0.01^{a}$	
G4	27.82.±2.12 ^b	0.24 ± 0.02^{b}	0.14 ± 0.01	1.28±0.17 ^b	0.16 ± 0.07^{b}	0.19 ± 0.04^{ab}	
G5	29.95±1.90 ^b	0.25±0.02 ^{ab}	0.13 ± 0.01	1.48 ± 0.18^{ab}	0.11 ± 0.01^{b}	0.16±0.01 ^b	
SIG	0.00	0.00	0.48	0.48	0.00	0.06	

G1: the negative control, G2: EMB, G3: EMB+ extract of Boswellia *serrate*, G4: EMB+ extract of Cinnamomum *zeylanicum* L, G5: EMB + powdered of the snail, sig: significant at p < 0.05, the values with different letters in each column showed a significant difference.

Antioxidants and oxidative markers

In approximately all living cells exposed to oxygen, SOD and CAT played a central role as antioxidant protection. In the current study, EMB-treated mice showed inhibition of SOD and CAT activity (Figure 2, 3). This may be due to a creation of ROS, namely H₂O₂ and superoxide anion. The accumulation of H₂O₂ and superoxide anion can activate some signaling pathways and lead to oxidative stress (Djordjevic et al., 2011). On the other side, EMB significantly amplified MDA concentration (Fig 1) at 60 days of exposure. The production of MDA is a biomarker that determined the level of oxidative stress (Pryor and Stanley, 1975). This increase in MDA agrees with the decrease in CAT and SOD when the mice are treated with EMB. MDA is an outcome of peroxidation for polyunsaturated fatty acid which resulted during degradation by ROS. This supports the incidence of cell toxic stress. Besides, the elevated level of MDA interacts

with DNA causing potentially mutagenic effects (Del Rio et al., 2005). Subsequently, DNA damage obtained herein was compatible with MDA results (Fig.1). These findings are parallel with those recorded by Zhu et al. (2013), who found that EMB inhibited the activity of SOD and increased the level of MDA. These results also agreed with some studies that indicated administration of EMB to mice or rats produces a significant increase of MDA while it decrees activities of CAT and SOD (El-Sheikh and Galal, 2015; Meligi and Hassan, 2017; Mossa et al., 2017). There are many substances that can ameliorate ROS induction by EMB. Abou-Zeid et al. (2018) used pumpkin seed oil with EMB with the mice that ameliorated the toxic effects including oxidative stress. This present study revealed that co-administration of EMB with BS, CZ and PS ameliorated the toxic effects of MDA, CAT, and SOD. Boswellic has anti-inflammatory and antioxidant properties. The intake of boswellic acid (an active compound of Boswellia species)

protected liver antioxidative and inflammatory injury induction by acetaminophen (Chen et al., 2016). Boswellic acid reduces the formation of MDA and increased SOD and CAT activity (Zhang et al., 2016). Oxidative stress is also inhibited by BS treatment with overall antioxidant potential which increased in the liver (Eltahir et al., 2019). MDA in these results decreased, meanwhile, CAT and SOD increased with co-administration with CZ. These results agreed with El Fadil et al. (2020) and Ojarudi et al. (2020). They stated that cinnamon extract had a defensive effect against oxidative stress produced by substances. The mechanism involves an increase in antioxidant enzymes (SOD and CAT) and a decrease in MDA levels. The defending action of cinnamon may be due to its inhibition effect on ROS generation during the production of flavonoids and phenolic compounds (Azab et al., 2011). These authors demonstrated indicate the importance of snail's products, which possibly lead to anti-inflammatory agents. Reviews also indicated that there is a lack of reports that have studied the molluscs anti-inflammatory activity (Ahmad et al., 2018). The treatment of EMB+PS increased the activity of antioxidants (CAT, SOD) and decreased MDA. This agreed with the previous literature about antioxidants and ROS relationship (Mossa, et al., 2017; Abou-Zeid, et al., 2018).



Figure 1. Effect of EMB alone or combined with natural compound on MDA level in mice.



Figure 2. Effect of EMB alone or combined with natural compound on SOD activity in mice



Figure 3. Effect of EMB alone or combined with natural compound on CAT activity in mice.

Biochemical analysis findings

Elevated plasma ALT and AST levels have been reported like a sign of liver cell damage (El-Shenawy and Abdel-Rahman, 1993). With a hepatocellular injury, the secretion of these enzymes increased in the blood. ALT and AST noticeably increase with EMB. This increase is sustaining the hypothesis that exposure to pesticide resulted in toxicity against biochemical liver (Hernández et al., 2013). The results diagrammatic in Figure (4) agreed with Khaldoun et al. (2017), who reported a markedly elevation of AST and ALT levels in the plasma of EMB-treated rats. These alterations of AST and ALT activity may be due to necrotic-changes of hepatic tissue that appear in histopathological examination (Figure. 7). Also, these results agreed with El-Sheikh and Galal (2015). Production of ROS caused damage to the diverse membrane components of the cell. This damage leads to infiltrate of cytoplasmic enzymes (Bagchi et al., 1995). Amelioration of the adverse effect of EMB on ALT and AST appears when BS, CZ and PS are treated with EMB (Figure 4). The findings in this respect are in agreement with that obtained by Chen et al. (2016), who found inhibition in serum levels of AST and ALT in boswellic acid-treated mice. In addition, histological obtained in this study data further supported the AST and ALT results. Eltahir et al. (2019) reported that BS markedly reduces CCl4-induced increases in ALT and AST levels. These results suggested that with its anti-inflammatory, antifibrotic and antioxidant characteristics, BS has hepatoprotective effects against toxic-substances cause liver injury. On the other hand, the treatment of EMB-treated mice with cinnamon ameliorates the adverse effect of EMB. Cinnamon reduces AST and ALT increase that induced by EMB. These results agreed with Hussain et al. (2019), who found that the mice treated with cinnamon markedly inhibited acetaminophendependent increases in the levels of AST and ALT. In addition, PS has a positive effect against the harmful effects of the insecticide on ALT and AST. It may be due to snail's products which have a wide array of healing properties. These therapeutic properties including antimicrobial, anticoagulant, immune modulating, wound healing, anticancer, antioxidant, antihypertensive, and antiinflammatory (Perdicalis et al., 2013).



Figure 4. Effect of EMB alone or combined with natural compounds on enzyme biomarkers in the liver of mice. ALT: Alanine aminotransferase, AST: Aspartate aminotransferase

Molecular analysis

The obtained qPCR results showed a significant upregulation in the expression levels of *Mgst1*, *Cyp2E1*,

Caspase3 and IL1b genes in the mice liver treated with EMB (G2) as compared to the negative control group (G1). This elevation expression was significantly downregulated following treatment with EMB+BS (G3), EMB+CZ (G4) and EMB+ PS (G5) (Fig.5). Microsomal glutathione Stransferase 1 (Mgst1) gene is responsible for the decrease of lipid hydroperoxides. It binds electrophiles with glutathione. The protein of this gene is a component of the outer membrane of the mitochondria and endoplasmic reticulum. It keeps these membranes of oxidative stress (Johansson et al., 2010). The cytochrome P450 (CYP450) is monooxygenases and catalyzes a lot of reactions implicated in the breakdown of toxic carcinogens. This enzyme contains subcategories like CYP2E1 subtype. CYP2E1 metabolizes both exogenous and endogenous substances (Lewis et al., 2003). Changes in the expression of CYP2E1 gene are involved in much pathology found by toxic substances (Zhou et al., 2009). This alteration may be due to its responsibility in procarcinogens metabolism and drugs as activator enzymes (Wang and Chou, 2010). These findings revealed that subchronic treatment of EMB to mice produced a significant upregulation in the expression levels of Mgst1and Cyp2E1genes in the liver. This elevated expression was significantly (P≤0.05) downregulated following the treatment with natural compounds (Figure.4). These results agree with Abou-Zeid et al. (2018), who reported that the Mgst1 and CYP2E1 genes expression markedly increased in the liver of emamectin-treated mice. In addition Cárcamo et al. (2011) and Cárcamo et al. (2017) found that the CYP gene expression was upregulated by the treatment with EMB in trout fish. As a result, this suggested that EMB changes the transcriptional process of proteins implicated in the distribution, metabolism and elimination of xenobiotics (Abou-Zeid et al. 2018). Similarly, the gene activity increased with treatment by many toxicants (Aniya et al. 2000). Mgst1 activation may be contributed to the depletion of glutathione and oxidized glutathione accumulation. The different measurements refer to activation of ROS related to the activation of Mgst1. This correlates with the results obtained herein where EMB activated the CYP2E1 expression and subsequently increased the Mgst1 expression. In accordance with these findings, the activation of CYP2E11, that causes extensive ROS release in a hepatoma cell line, was shown to be improved the expression and activation of Mgst1 (Marí and Cederbaum, 2001). Caspase-3 is the major apoptosis effector protein that is induced by a caspase initiator as caspase-9. The results indicated that EMB-induced cell apoptosis through activation of both caspase-9 and caspase-3 decay intracellular proteins and carry out the program of cell death (Zhang et al., 2017). These findings revealed that subchronic treatment of EMB to mice caused a markedly upregulation in the caspase3 gene expression levels in the liver. These results agreed with Azoz et al. (2020) who observed that the caspase-3 gene expression obviously

increased after administrated rats with EMB. The apoptosis is activated by EMB linked to ROS generation. In turn, ROS triggers the activation pathway of the mitochondrialdependent intrinsic. It leads to disruption function of mitochondria and consequent potential mitochondrial membrane breakdown to be released cytochrome-c (Zhang et al., 2017). Cytokines are considered essential players in disorders associated with inflammation throughout the body. The quantity of inflammation affects the long term result of liver disease. The pro-inflammatory cytokines Interleukin (IL)-1 play a vital role in several stages of liver diseases. (Niederreiter and Tilg 2018). On the other hand, insecticides are linked with direct or indirect alteration of vital inflammatory activation and immune mechanisms (Miller et al., 2009). Celik-Ozenci et al. (2011) and Banerjee et al. (2001) established the poisoning of pesticides resulted in the creation of free radicals in organisms and inducing oxidative stress. The free radicals increasing perhaps make modifications in the cell structures and affect the immune function (Liu et al., 2014). Exposure to many insecticides accelerates the formation of IL-1b cytokines (Duzguner and Erdogan, 2010). Liu et al. (2014) reported that the creation of cytokines perhaps acts like a kind of compensatory response after undesirable stress effects on an organism. Therefore, IL-1b cytokine is a desirable indicator for studies belong to immune-suppression and related to poisons. The results revealed that mice treated with EMB produces a significant upregulation in the *IL1b* gene expression levels in the liver. These outcomes are in parallel agreed with that reported by Liu et al., (2014), who found that avermectin caused immune suppression by increasing the *IL-1* β mRNA levels in the pigeon. Also, Duzguner and Erdogan, (2014), found that imidacloprid stimulated IL-1b expressions in the liver. On the other hand, treatment with natural compounds led to a downregulation the expression of genes as a result of reducing the harmful effects of the insecticide. Boswellic acid administration inhibited the elevation of cytokines and CYP2E1. Consequently, it lowered ROS production and reduced oxidative stress in the mice liver (Chen et al., 2016). Gayathri et al. (2007) demonstrated that extract of Boswellia serrata downregulated IL-1 β cytokines in peripheral blood mononuclear cells. The cinnamon extract has an immune stimulant outcome on the proliferation of human lymphocytes and *IL-1\beta* construction by monocytes (Shan *et* al., 1999). It regulates immune function via regulating pro and anti-inflammatory mediators and the gene expression in macrophages (Cao et al., 2008). These explain the cinnamon role in adjustable gene expression in EMB treated mice. The data revealed that treatment with snail powder caused a significant downregulation in the expression levels of studied genes in the liver to be compared with the positive control. This agreed with Sarkar et al. (2015), who support the anti-inflammatory function of the Indian freshwater snail.



Figure 5. Relative expression of *Mgst1*, *CYP2E1*, *Caspase 3* and *IL1b* (compar to β-actin as internal control) genes in the liver of mice treated with EMB alone or combined with natural compounds.

Comet assay

Emamectin administration to mice induced a marked increase in liver DNA fragmentation (Figure 6 and Table 6).

These results agreed with Zhang et al. (2017) and Yun et al. (2017), who detected apoptosis in cells of EMB human liver exposure by enhanced caspase-3 activities and increased DNA fragmentation. This agreed with Azoz et al. (2020), who found that EMB oral intake by rats caused a markedly increase in DNA damage. Insecticides exposure is reported to produce DNA break that leads to the process involved in cells genotoxic effect (Dusinska and Collins 2008; Mater et al., 2014). The fragmentation of nuclei and condensation of chromatin were noticed resulting from EMB exposure in Sf-9 cells (Wu et al., 2016). On the other, treatment with BS reduce the tail length, tail DNA%, and tail moment. This agreed with Rajabian et al. (2016), who found that DNA damage in BS-treated cells with glutamate was markedly decreased if compared with glutamate-treated cells. Moreover, consistent with these results B. serrata also inhibited the oxygen radicals (Ammon, 2010). Alcohol and aqueous extract of BS has high activity in scavenging free radicals (Sharma et al., 2011; Azemi et al., 2012). Regarding cinnamon extract, Sağlam et al. (2012) demonstrated that the length of comet tail was decreased in the diabetic group treated with cinnamon. Karadağlı (2014) showed that cinnamon is a protective agent in inhibiting damage induced by oxidative stress. The ability of cinnamon inhibited DNA damage may be due to its containing of high flavonoid and polyphenol compounds (Kumar et al., 2012).

 Table 6. Comet parameters in the liver of mice treated with EMB alone or combined with natural compounds.

Group	Tailed	Untailed	Tails	Tail	Tail
	%	%	length µm	DNA%	moment
G1	1.75	98.25	1.45±0.53 ^d	1.36	1.97 ± 0.69^{d}
G2	32	68	9.63±1.27 ^a	7.41	71.36±9.10 ^a
G3	18	82	7.44±0.93 ^b	5.60	41.66±7.41 ^b
G4	14	86	6.21±0.90 °	4.22	26.21±3.57°
G5	20	80	8.03±1.11 ^b	6.16	49.46±7.14 ^b

Different superscript letters in the same column of tail length showed significance difference at P< 0.05





Fig. 6. Representative images of the cells assayed by the comet assay. Where G1 = negative control; G2 = mice treated with EMB;G3= mice treated with EMB+BS; G4 = mice treated with EMB+CZ; G5 mice treated with EMB+ PS.

Histopathological findings

The most histological effects related to the treated mice with EMB are shown in Figure.7 (G2 a and b). The treatment *Boswellia* with EMB enhanced the architecture of liver. It showed slight activation of kupffer cells, slight infiltration of lymphocytes, karyolysis of some nuclei and normal central vein (Figure 7, G3). The same results appeared with the treatment by methyl extract of *Cinnamonum*. It revealed hepatocytes recovering with minimal vacuoles. Also, it can be seen that the normal

Sherifa F. M. Dawoud et al

central vein and hepatocytes (Figure 6 G4. The treatment with PS decreased the side effects induced by EMB in mice liver (Figure 6 G5).

The results belong to histopathological studies with EMB treatment agreed with El-Sheikh and Galal (2015), Khaldoun et al. (2017) and Abou-Zeid et al. (2018), who found that emamectin administration produced pathological changes in the liver. These changes include blood vessel congestion with infiltration of lymphocytes, necrosis with hepatic parenchyma infiltration, also, dilated sinusoids. The EMB-side effects caused by the production of ROS that induced many changes in the cell. This clearly appears in icreasing MDA as seen in Figure 1. ROS generated by EMB can damage membrane components of the cell and lead to the leakage of cytoplasmic enzymes (Bagchi et al., 1995). This appeared in ALT and AST activites (Figure 4). Also the oxiditive stress can lead to apoptosis (Temiz, 2020). This cleary appeared in gene expression of caspas3 (Figure. 5). Khaldoun et al. (2017) suggested that EMB toxicity is possibly caused by oxidative stress. The BS treatment combined with EMB leads to recovery and return to the normal appearance of hepatocytes and central vein. The most histological effects showed slight lymphocyte infiltration and karyolysis of some nuclei (Figure 6, G3). These results agreed with Eltahir et al. (2019), who reported that BS has antifibrotic characteristics, anti-inflammatory and antioxidant against CCl4 that induced degeneration in the liver. Also, it inhibits the biosynthesis of leukotrienes that considered pro-inflammatory mediators (Zhang et al., 2013). Leukotrienes increased cell permeability. Also, boswellic acid inhibits transcription factor NF-KB. It is a vital downstream mediator for cytokines during inflammation (Cuaz-Pérolin et al., 2008) that can also concurrently reduced oxidative stress (Umar et al., 2014). Cinnamon reduced the side effect of EMP in liver tissue. It removed the histopathological changes through anti-oxidant activity (Hussain et al., 2019). The cinnamon also has antiinflammatory activity and free radicals-scavenge ability. So, a supplemented diet with cinnamon increased the protection against anti-toxic exposure (Bellassoued et al., 2019). Ziconotide, first isolated from the snail Conus magus venom significantly blocks N-type voltage-gated calcium channels (Schroeder et al., 2004). It is effective for the healing chronic pain. There is no sufficient studies took place about snail powdered and its role in eliminating the damage in the liver induced by toxic substances.



Figure 7. Histopathological changes

CONCLUSION

It can be concluded that EMB induced changes in the gene expression of *Mgst1*, *CYP2E*, *caspase-3* and *IL-1β*. Also, it is induced DNA fragmentation with increasing MDA, AST, and ALT. In contrast, it decreased both CAT and SOD levels. The treatment with natural extracts can remove the side effects of EMB. The findings belong to EMB and natural compounds are new in this filed beside there is no sufficient studies on the PS belong to this topic. So it can be used these natural materials to avoid the side effects of insecticide and other toxin substances.

REFERENCES

- Abd-Allah, S.H., Shalaby, S.M., Abd-Elbary, E., Saleh, A.A. and El-Magd, M.A. (2015). Human peripheral blood CD34+ cells attenuate oleic acid-induced acute lung injury in rats. Cytotherapy.17:443-453.ISSN 14772566,14653249 https://doi.org/10.1016/j.jcyt.2014.11.002.
- Abou-Zeid, S.M., Abu Bakr, H.O., Mohamed, M.M. and El-Bahrawyd, A. (2018). Ameliorative effect of pumpkin seed oil against emamectin induced toxicity in mice. Biomedicine & Pharmacotherapy .98:242–251. ISSN07533322. https://doi.org/ 10. 1016/ j.biopha.2017.12.040.
- Aebi, H.(1984). Catalase in vitro. Methods Enzymol.105: 121–126. ISSN 00766879,15577988 https://doi.org/ 10.1016/S0076-6879, 84)05016-3.
- Ahmad, T.B., Liu, L., Kotiw, M. and Benkendorff, K. (2018). Review of anti-inflammatory, immune-modulatory and wound healinproperties of molluscs. Journal of Ethnopharmacology. 210:156–178. ISSN03788741, 18727573

.http://dx.doi.org/10.1016/j.jep.2017.08.008.

- Ammon, H.(2010).Modulation of the immune system by Boswellia serrata extracts and boswellic acids. Phytomedicine.17:862–867. ISSN 1572557X. doi: 10.1016/j.phymed.2010.03.003.
- Aniya, Y., Ohtani, I.I., Higa, T., Miyagi, C., Gibo, H., Shimabukuro, M., Nakanishi, H. and Taira, J.(2000). Dimerumic acid as an antioxidant of the mold, Monascusanka, Free Radic. Biol. Med. 28: 999–1004. ISSN-08915849, 18734596 doi: 10.1016/s0891-5849(00)00188-x.
- Awoke,D.S. E. and Joshi, R. K. (2021). Analysis of major components of essential oils of Boswellia species by GC-MS. American Journal of Essential Oils and Natural Products, 9(1), 48-54.
- Azab, K.S., Mostafa, A.H.A., Ali, E.M.M. and Abdel-Aziz, M.A.S.(2011). Cinnamon extract ameliorates ionizing radiation-induced cellular injury in rats. Ecotoxicology and Environmental Safety.74: 2324 2329.ISSN 01476513, 10902414.doi: 10.1016 /j. ecoenv .2011.06.016. Epub 2011 Jul 22.
- Azemi, M.E., Namjoyan, F., Khodayar, M.J., Ahmadpour, F., DarvishPadok, A. and Panahi, M.(2012). The antioxidant capacity and anti-diabetic effect of *Boswellia serrata Triana* and *Planch* aqueous extract in fertile female diabetic rats and the possible effects on reproduction and histological changes in the liver and kidneys. Jundishapur J Nat Pharm Prod. 7: 168– 675. ISSN 22287876, 17357780. DOI: 10.5812 /jjnpp.6755.
- Azoz, A, Ibrahim KA, Abdel Kader IY, Tawfik A. Tracking of Apoptosis Induced by Emamectin Benzoate in Fetuses of Hypothyroid Rats.Int. J. Pharm. Sci. Rev.Res. 13:81-89. ISSN0976044X. https://www. researchgate.net/publication/343167464.

- Bagchi, D., Bagchi, M., Hassoun, E.A. and Stohs, S.J.(1995). In vitro andin vivo generation of reactive oxygen species, DNA damage and lactate dehydrogenase leakage by selected pesticides. Toxicology.104: 129– 140. ISSN 0300483X, 18793185. doi: 10.1016/0300-483x(95)03156-a.
- Banerjee, B.D., Seth V. and Ahmed, R.S. Pesticide-induced oxidative stress: perspective and trends. Rev. Environ. Health. 16 (1): 1–40.doi: 10.1515/reveh.2001.16.1.1.
- Bellassoued, K., Ghrab, F., Hamed, H., Kallel, R., Pelt, J.V., Lahyani, A., Ayadi, F.M. and El Feki, A. (2019).Protective effect of *CinnamomunzeylanicumL*. bark essential oil, on hepatic and renal toxicity induced by CCl4 in rats.Applied Physiology, Nutrition, and Metabolism. 44(6):606-618. doi: 10.1139/apnm-2018-0246. Epub 2019 Apr 17.
- Bryman, A.; Cramer, D. (2011). Quantitative Data Analysis with IBM SPSS 17, 18 and 19: A Guide for Social Scientists. New York: Routledge. ISBN 978-0-415-57918-6
- Cao, H., Urban, J.F. and Anderson, R.A.(2008). Cinnamon polyphenol extract affects immune responses by regulating anti- and pro-inflammatory and glucose transporter gene expression in mouse macrophages. J Nutr. 138:833-40.ISSN 15416100, 00223166. DOI: 10.1093/jn/138.5.833.
- Cárcamo, J.G., Aguilar, M.N., Barrientos, C.A., Constanza, F., Carreño, C.F., Quezada, C.A., Bustos, C., Manríquez, R.A., Avendaño, H. R. and Yañez, A.J.(2011). Effect of emamectin benzoate on transcriptional expression of cytochromes P450 and the multidrug transporters (Pgp and MRP1) in rainbow trout (Oncorhynchusmykiss) and the sea lice Caligusrogercresseyi.Aquaculture. 321:207– 215. ISSN 00448486.DOI: 10.1016/ j.aquaculture .2011;09.012.
- Cárcamo, J.G., Aguilar, M.N., Carreño, C.F., Vera, T., Arias-Darraz, L., Figueroa, J.E., Romero, A.P., Alvarez, M. and Yañez, A.J.(2017). Consecutive emamectin benzoate and deltamethrin treatments affect the expressions and activities of detoxification enzymes in the rainbow trout (Oncorhynchusmykiss), Comp. Biochem. Physiol. C: Toxicol. Pharmacol. 191: 129– 137. ISSN 15320456, 18781659 doi: 10.1016/j.cbpc. 2016.10.004. Epub 2016 Oct 17.
- Celik-Ozenci, C., Tasatargil, A., Tekcan, M., Sati, L., Gungor, E., Isbir, M. and Demir, R.(2011). Effects of abamectin exposure on male fertility in rats: potential role of oxidative stress-mediated poly (ADP-ribose) polymerase (PARP) activation. Regul. Toxicol. Pharm.61 (3): 310– 317.ISSN02732300,10960295. <u>https://doi.org/ 10.1016 /j</u> .yrtph.2011.09.001.
- Chen, L.C., Hub, L.H. and Yinc, M.C.(2016).Alleviative effects from boswellic acid on acetaminophen-induced hepatic injury.Bio Medicine. 6 (2):12-19.ISSN 2211-8039.10.7603/s40681-016-0009-1.
- Cuaz-Pérolin, C., Billiet, L., Baugé, E., Copin, C., Scott-Algara, D., Genze, F., Büchele, B., Syrovets, T., Simmet, T. and Rouis, M.(2008).Anti-inflammatory and antiatherogenic effects of the NF-kappaB inhibitor acetyl-11-keto-beta-boswellic acid in LPS-challenged ApoE-/- mice. ArteriosclerThrombVasc Biol. 28: 272-277.
- Del Rio, D., Stewart, A.J. and Pellegrini, N.(2005). A review of recentstudies on malondialdehyde as toxic molecule and biologicalmarker of oxidative stress. Nutr. Metabol. Cardiovasc. Dis. 15:316–328. ISSN 15903729, 09394753.doi: 10.1016/j. numecd. 2005. 05.003.
- Djordjevic, J., Djordjevic, A., Adzic, M., Elakovi'c, I., Mati'c, G. and Radojcic, M.B.(2011). Fluoxetine affects antioxidant systemand promotes apoptotic signaling in wistar rat liver. Eur. J.Pharmacol.659: 61–66. ISSN 00142999, 18790712 .doi: 10.1016/j.ejphar.2011.03.003. Epub 2011 Mar 21.

- Dusinska, M. and Collins, A.R.(2008). The comet assay in human biomonitoring: gene environment interactions. Mutagenesis. 23: 191–205. ISSN 02678357, 14643804.doi: 10.1093/mutage/gen007.
- Duzguner, V. and Erdogan, S.(2010). Acute oxidant and inflammatory effects of imidacloprid on the mammalian central nervous system and liver in rats.Pest.Biochem. Physiol. 97: 13–18. ISSN 10959939, 00483575. https://doi.org/ 10.1016/ j.pestbp .2009.11.008
- Duzguner, V. and Erdogan, S.(2014).Chronic exposure to imidacloprid induces inflammation and oxidative stress in the liver & central nervous system of rats.Pesticide Biochemistry and Physiology.;104:58-64.http://dx.doi.org/10.1016/j.pestbp.2012.06.011.
- Eidi, A., Mortazavi, P., Bazargan, M. and Zaringhalam J.(2012). Hepatoprotective activity of cinnamon ethanolic extract against CCL4-induced liver injury in rats.EXCLI Journal. 2012;11: 495-507.ISSN 16112156. DOI: 10.17877/DE290R-4957.
- El Fadil, H.A., Moustafa, A., Khalifa, H., Hossam, A. and Behairy, A.(2020). Cinnamon Extract Ameliorates Liver Damage And Oxidative Stress Induced By Paracetamol In Male Rats. Damanhour journal of veterinary sciences. 3 (2): 14-20.
- El-Bialy, B.E., AbdEldaim, M.A., Hassan, A. and Abdel-Daim, M.M.(2020). Ginseng aqueous extract ameliorates lambda- cyhalothrinacetamiprid insecticide mixture for hepatorenal toxicity in rats: role of oxidative stress-mediated proinflammatory and proapoptotic protein expressions. Environ Toxicol. 35(2):124–135.ISSN15227278, 15204081.doi: 10.1002/tox.22848. Epub 2019 Sep 30.
- Eldamaty, S.E., Elbasiouny,H., Elmoslemany, A.M. Abd El-Maoula, L.M., El-Desoky, O.I., Rehan,M. El Moneim, D.A. and Zedan, A. (2021). Protective Effect of Wheat and Barley Grass Against the Acute Toxicological Effects of the Concurrent Administration of Excessive Heavy Metals in Drinking Water on the Rats Liver and Brain. Appl. Sci. 11, 50-59.
- El-Sheikh, E. and Galal, A.A.(2015). Toxic effects of sub-chronic exposure of male albino rats to emamectin benzoate and possible ameliorative role of Foeniculumvulgare essential oil. Environmental toxicology and Pharmacology. 39: 1177– 1188.ISSN13826689.doi: 10.1016/ j.etap. 2015. 04.008.
- El-Shenawy, N.S. and Abdel-Rahman, M.S.(1993). The mechanism of chloroform toxicity in isolated rat hepatocytes. Toxicology Letters. 69: 77–85. ISSN 03784274, 18793169. https://doi.org/10.1016/0378-4274(93)90148-Q.
- Eltahir, H.M., Fawzy, M.A., Mohamed, E.M., Alrehany, M.A., Shehata, AM.and Abouzied, M.M.(2019). Antioxidant, antiinflammatory and anti-fibrotic effects of Boswellia serrata gum resin in CCl4-induced hepatotoxicity. Experimental and therapeutic medicine. 19(2):1313-1321.ISSN17921015, 17920981. DOI:10.3892/etm.2019.8353.
- Gayathri, B.N., Manjula, N., Vinaykumar, K.S., Lakshmi, B.S., Balakrishnan, A.(2007). Apure compound from Boswelliaserrata extract exhibits anti-inflammatory property in human PBMCs and mouse macrophages through inhibition of TNFα, IL-1β, NO and MAP kinases.International Immunopharmacology.7:473– 482. ISSN 18781705, 15675769. doi:10.1016/j.intimp .2006.12.003.
- Hadi, M.Y. and Hameed, I, H. (2017). Uses of Gas Chromatography-Mass Spectrometry (GC-MS) Technique for Analysis of Bioactive Chemical Compounds of Lepidium sativum: A Review. Research J. Pharm. and Tech. 10(11):4039-4042. DOI: 10.5958/0974-360X.2017.00732.6.

- Hayes, W.J. and Laws, E.R.(2013). Handbook of pesticide toxicology, classes of pesticides. Jr. Elsevier. 3: 1451–1453.
- Hernández, A.F., Gil, F., Lacasa na, M., Rodríguez-Barranco, M., Tsatsakis, A.M., Requena, M., Parrón, T. and Alarcón, R. (2013).Pesticide exposure and genetic variation inxenobiotic-metabolizing enzymes interact to induce biochemical liver damage. Food Chem. Toxicol. 61:144–151. ISSN 18736351, 02786915.doi: 10.1016/j.fct.2013.05.012. Epub 2013 May 18.
- Hussain, Z., Khan, J.A., Arshad, A., Asif, P., Rashida, H. and Arshad, M.I.(2019).Protective effects of Cinnamomumzeylanicum L. (Darchini) in acetaminophen- induced oxidative stress, hepatotoxicity and nephrotoxicity in mouse model. Biomedicine & Pharmacotherapy. 109 : 2285–2292.I SSN .07533322 https://doi.org/10.1016/j.biopha.2018.11.123.
- Johansson, K., Järvliden, J., Gogvadze, V. and Morgenstern, R.(2010). Multiple roles of microsomal glutathione transferase 1 in cellular protection: a mechanistic study. Free Radic. Biol. Med. 49: 1638–1645. ISSN 08915849,18734596. doi: 10.1016/ j.freerad biomed. 2010.08.013.
- Kalairasan, A., Kumar, P. and Ahmed, J.S. (2011). GC/MS determination of bioactive components of Bulbophyllum kaitense. Reichib leaves estern ghats in India. NY Sci 4(10):46–49.
- Karadağlı, S.S.(2014). Investigation of the protective effect of cinnamomum cassia bark extract against H₂O₂-induced oxidative DNA damage in human peripheral blood lymphocytes and antioxidant activity. Marmara Pharmaceutical Journal. 18(1): 43-48. ISSN 13090801.DOI: 10.12991/mpj.201414125.
- Khaldoun, O,H., Richeval, C., Lebaili, N., Zerrouki-Daoudi, N., Baha, M., Djennas, N. and Allorge, D.(2017). Ameliorative effect of vitamin C against hepatotoxicity induced by emamectin benzoate in rats. Human and experimental toxicology. 36 (7):709–717. ISSN 09603271. DOI: 10.1177/0960327116661022.
- Khaldoun, O.H., Allorgec, D., Richevalc, C., Lhermittec, M. and Djenase, N.(2015).Emamectin benzoate (Proclaim®) mediates biochemical changes and histopathological damage in the kidney of male Wistarrats (*Rattusnorvegicus*). Toxicologie Analytique&Clinique.27:72-80. ISSN 23520078.http://dx.doi.org/10.1016/j.toxac.2014.11.002.
- Khamis, A.A.A., Ali, E.M.M., El-Moneim, M.A.A., Abd-Alhaseeb, M.M., El-Magd, M.A. and Salim, E.I.(2018). Hesperidin, piperine and bee venom synergistically potentiate the anticancer effect of tamoxifen against breast cancer cells, Biomed. Pharmacother105:1335-1343. ISSN 07533322 .DOI: 10.1016/j.biopha.2018.06.105.
- Kumar, M., Sharma, V.L. and Sehgal, A. and Jain, M.(2012). Protective effects of green and white tea against benzo (a) pyrene induced oxidative stress and DNA damage in murine model. Nutrition and cancer. 64(2): 300-306. ISSN 01635581,15327914.doi: 10.1080/01635581.2012.648300. Epub 2012 Jan 13.
- Leal, M.C., Madeira, C., Brandao, C.A., Puga, J. and Calado, R.(2012). Bioprospecting of marine invertebrates for new natural products - a chemical and zoogeographical perspective.Molecules. 17 (8): 9842–9854. ISSN 14203049.DOI: 10.3390/ molecules 17089842.
- Lewis, D.F., Lake, B.G, Bird, M.G., Loizou, G.D., Dickins, M. and Goldfarb, P.S.(2003).Homology modelling of human CYP2E1 based on the CYP2C5 crystal structure: investigation of enzyme-substrate and enzyme-inhibitor interactions. Toxicol. Vitro. 17: 93–105. ISSN18793177, 08872333.doi: 10.1016/s0887-2333(02)00098-x.
- Liu, C., Li, M., Cao, Y., Qu, J., Zhang, Z., Xu, S. and ShuLi, S.(2014).Effects of avermectin on immune function and oxidative stress in the pigeon spleen.Chemico-Biological Interactions. 210: 43–50. ISSN00092797. http://dx.doi.org/10.1016/j.cbi.2013.12.015.

- Mansour, S.A. and Mossa, A.T.H.(2010).Oxidative damage,biochemical and histopathological alterations in rats exposedtochlorpyrifos and the antioxidant role of zinc. Pest.Biochem.Physiol.96: 14–23. ISSN 10959939- 00483575.https ://doi.org/ 10.1016/ j.pestbp. 2009.08.008
- Marí, M. and Cederbaum, A.I.(2001).Induction of catalase, alpha, and microsomal glutathione S-transferase in CYP2E1 overexpressing HepG2 cells and protection against shortterm oxidative stress, Hepatology. 33: 652–661. ISSN16000641 ,01688278 <u>https://doi.org/ 10.1053/jhep.2001.22521</u>.
- Mark, D.J (2018). Asthma.Integrative Medicine (Fourth Edition). 288-299.e2.ISBN 9780323358682. https://doi.org/10.1016/B978-0-323-35868-2.00029-3(http://www.sciencedirect.com/science/article/ pii/B9780323358682000293). Chapter 29 -, Editor(s): David Rakel, Elsevier.
- Mater, N., Geret, F., Castillo, L., Faucet-Marquis, V., Albasi, C., Pfohl-Leszkowicz, A.(2014). In vitro tests aiding ecological risk assessment of ciprofloxacin, tamoxifen and Cyclophosphamide in range of concentrations released in hospital waste-water and surface water. Environ. Int. 191– 200. ISSN18736750, 01604120.<u>https:// doi.org/ 10.1016/ i.envint.2013.11.011</u>.
- Meligi, N.M.and Hassan, H.F.(2017).Protective effects of Eruca sativa (rocket) on abamectin insecticide toxicity in male albino rats. Environ. Sci. Pollut. Res. Int. 24:9702–9712.ISSN09441344, 16147499.DOI 10.1007/s11356-017-8671-8.
- Miller, R.L., James-Kracke, M., Sun, G.Y. and Sun, A.Y.(2009). Oxidative and inflammatory pathways in Parkinson's disease. Neurochemistry 2009; 34: 55–65. ISSN 18729754, 01970186.doi: 10.1007/s11064-008-9656-2.
- Morgana, A.M., El-Ballal, S.S., El-Bialy, B.E. and EL-Borai, N.B.(2014). Studies on the potential protective effect of cinnamon against bisphenol A- and octylphenolinduced oxidative stress in male albino rats.Toxicology Reports. 1: 92–101.ISSN 22147500. doi: 10.1016/j. toxrep. 2014.04.003. e Collection 2014.
- Mossa, A.H., Abdel Rasoul, M.A. and Mohafrash, S.M. (2017) Lactational exposure to abamectin induced mortality and adverse biochemical and histopathological effects in suckling pup. Environ. Sci. Pollut. Res. Int. 24:10150–10165.ISSN09441344, 16147499.doi: 10.1007/s11356-017-8600-x.
- National Registration Authority for Agricultural and Veterinary Chemicals 2011, Ag Manual: The Requirements Manual for Agricultural Chemicals, NRA, Canberra, Australia.
- Niederreiter, L. and Tilg, H. (2018).Cytokines and fatty liver diseases. Liver Research 2 : 14-20. ISSN 25425684 https://doi.org/10.1016/j.livres.2018.03.003.
- Nishikimi, M., Appaji, N. and Yagi, K. (1972). The occurrence of superoxide anion in the reaction of reduced phenazinemethosulfate and molecular oxygen.Biochem.Biophys. Res. Commun. 46:849– 854. ISSN0006291X, 10902104. <u>https://doi.org/</u> 10.1016/ S0006-291X (72) 80218-3.
- Ohkawa, H., Ohishi, W. and Yagi, K. (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal. Biochem. 95: 351–358. ISSN 10960309, 00032697 doi: 10.1016/0003-2697 (79)90738-3.
- Ojarudi, M., Moradi, A., Hajihosseini, R., Mazani, M. and Rezagholizadeh, L. (2020). Hepatoprotective and antioxidant activities of combination of *Cinnamonum zeylanicum* and *Zingiberofficinale* in CCl4-intoxicated Rats.Journal of Kerman University of Medical Sciences. 27 (1): 1-13.ISSN 20082843,10239510. doi10.22062/jkmu.2020.89591.

- Perdicalis, S., Vlachogianni, T. and Vlavanidis, A.(2013). Bioactive natural substances from marine sponges: new developments and prospects for future pharmaceuticals. Nat. Prod. Chem. Res. 1 (3): 1–8. DOI: 10.4172/2329-6836.1000115.
- Prasanna, B. and Anand, A.V.(2019). *Cinnamon species: In vivo* anti-oxidant activity of ethanolic extracts of *Cinnamon zeylanicum and Cinnamon cassicae* barks. Pharmacog J.11(2):245-247.
- Pryor, W.A. and Stanley, J.P.(1975). Suggested mechanism for theproduction of malonaldehyde during the autoxidation of polyunsaturated fatty acids.Non enzymatic production of prostaglandin endoperoxides during autoxidation. J. Org.Chem. 40: 3615–3617. ISSN 15206904, 00223263. <u>https://doi.org/ 10.1021/</u> jo00912a038.
- Rajabian, A., Boroushaki, M.T., Hayatdavoudi, P. and Sadeghnia, H.R.(2016).boswellia serrata protects against glutamate-induced oxidative stress and apoptosis in pc12 and n2a cells. DNA and cell biology. 35(11): 1–14. ISSN 10445498. DOI: 10.1089/dna.2016.3332.
- Rehana, B.H. and Nagarajan, N. (2013). GC–MS determination of bioactive components of Wedelia chinensis (Osbeck) Merrill. J Chem Pharm Res. 5(4):279–285.
- Reitman, S. and Frankle, S. (1957). Coloremetric method for determination of serum transaminase activity. Am. J. Clin.Path. 28:56–68. ISSN19437722, 00029173 doi: 10.1093/ajcp/28.1.56.
- Roberts, E.(1984). GABA neurons in the mammalian central nervous system: model for a minimal basic neural unit.Neurosci. Lett.47: 195–200.ISSN 18727972 ,03043940.https://doi.org /10.1016/03043940(84)90513-5.
- Sağlam, O., Değirmenci, I., Üstüner, M.C.and Güneş, H.V.(2012). The protective effects of cinnamon and sugar tea extract on diabetic rats with interrelationships between oxidative stress and DNA damage. African journal of pharmacy and pharmacology.6(43): 3012-3017. ISSN 1996-0816. DOI: 10.5897/AJPP12.539.
- Sarkar, A., Gomes, A. and Gomes, A. (2015). Anti-osteoarthritis, antinociception, anti-inflammatory activities of isolated fraction of flesh extract Viviparous bengalensis in experimental model, Int. J. Curr. Res. Acad. Rev. 3 (6): 66–87.ISSN 09755241, 22312196.www.ijcrar.com.
- Schroeder, C.I., Smythe, M.L. and Lewis, R.J.(2004).Development of small molecules that mimic the binding of omega-conotoxins at the N-type voltage-gated calcium channel, Mol. Divers. 8 (2004) 127–134. ISSN 1573501X,13811991doi: 10.1023/b:modi.0000025656.79632.86.
- Shan, B.E., Yoshida, Y., Sugiura, T. and Yamashita,U.(1999).Stimulating activity of Chinese medicinal herbs on human lymphocytes in vitro.Int J Immunopharmacol. 21: 149-159.ISSN 01623109.doi: 10.1016/s0192-0561(98)00074-5.
- Shareef, H. K., Muhammed, H. J., Hussein, H. M., & Hameed, I. H. (2016). Antibacterial effect of ginger (Zingiber officinale) roscoe and bioactive chemical analysis using gas chromatography mass spectrum. *Oriental Journal of Chemistry*, 32(2), 20-40.
- Sharma, A., Upadhyay, J., Jain, A., Kharya, M.D., Namdeo, A. and Mahadik, K.R.(2011). Antioxidant activity of aqueous extract of Boswellia serrata. J Chem Bio PhySci 2011;1:60–71. ISSN 2249–1929.
- Shen, Y., Jia, L., Honma, N., Hosono, T., Ariga, T. and Seki, T.(2012). Beneficial effects of cinnamon the metabolic syndrome, inflammation pain and mechanism underlying these effects a review.Journal of Traditional and Complementary Medicine. 2(1): 27-32.ISSN22254110. doi: 10.1016/s2225-4110 (16) 30067-0.

- Shihabudeen, M.S., Priscilla, H. and Thirumurugan, K.(2011).Cinnamon extract inhibits glucosidase activity and dampens postprandial glucose excursion in diabetic rats. Nutrition & Metabolism. 8 (1):46.ISSN17437075. DOI: 10.1186/1743-7075-8-46.
- Sreekumar, V.T., Ramesh, V. and Vijaykumar, R. (2014). Study on ethanolic extract of Pitchavari: a native medicinal rice from southern peninsular India. Int J Pharm Sci Rev Res 25(2):95–99
- Suvarna, S.K., Layton, C. and Bancroft, J.D.(2013). Bancroft's theoryand practice of histological techniques, 7th Ed. Elsevier, Churchill Living stone, England.https: //www. elsevier. com/books/bancrofts-theory- and- practice-ofhistological-techniques-e-book/ suvarna/ 978-0-7020-5032 -9. eBook ISBN: 978070 2050329.
- Temiz, O.(2020).The potential of emamectin benzoate to induce kidney DNA oxidation, heat shock protein levels and apoptosis in male mice. Acta scientific pharmaceutical sciences. 4. (2): 01-05. DOI: 10.31080/ASPS.2020.04.484.
- Umar, S., Umar, K., Sarwar, A.H., Khan, A., Ahmad, N., Ahmad, S., Katiyar, C.K., Husain, S.A. and Khan, H.A.(2014).*Boswellia serrata* extract attenuates inflammatory mediators and oxidative stress in collagen induced arthritis. Phytomedicine. 21:847-856.ISSN09447113, 1618095X. doi: 10.1016/j.phymed.2014.02.001. Epub 2014 Mar 22.
- Wang, J.F. and Chou, K.C.(2010). Molecular modeling of cytochrome P450 and drug metabolism, Curr.Drug Metab. 11 : 342–346. ISSN 18755453, 13892002 doi: 10.2174/138920010791514180.
- Wolterink, G., Kesteren, P. and Mc-Gregor, D.(2011). Emamectin Benzoate. JMPR. 2011;211–252.
- Wu, X., Zhang, L., Yang, C., Zong, M., Huang, Q. and Tao, L. (2016). Detection on emamectin benzoate-induced apoptosis and DNA damage in *Spodopterafrugiperda*Sf-9 cell line. Pesticide Biochemistry and Physiology. 126: 6-12.ISSN10959939, 00483575 http://dx.doi.org/ 10.1016/ j.pestbp.2015.06.009.
- Yassin, N.A.Z., El-Shenawy, S.M.A., Mahdy, K.A., Gouda, N.A.M., Abd El- Marrie, F.H., Farrag, A.H. and Ibrahim, B.M.M. (2013). Effect of *Boswellia serrata* on Alzheimer's disease induced in rats. Journal of the Arab Society for Medical. 8:1–11. DOI: 10.7123/ 01. JASMR. 0000429323. 25743.cc.

- Yazdanpanahi, N., Behbahani, M. and Yektaeian A.(2014).Effect of *Boswellia Thurifera* Gum Methanol Extract on Cytotoxicity and *P53* Gene Expression in Human Breast Cancer Cell Line. Iranian Journal of Pharmaceutical Research. 2014;13 (2): 719-724.ISSN 17350328, 17266890 doi10.22037 /ijpr.2014.1507.
- Yun, X., Rao, W., Xiao, C. and Huang Q.(2017). Apoptosis of leukemia K562 and Molt-4 cells induced by emamectin benzoate involving mitochondrial membrane potential loss and intracellular Ca²⁺ modulation. Environ. Toxicol.Pharmacol. 52:280– 287. ISSN 13826689.doi: 10.1016/j.etap.2017.04.013. Epub 2017 Apr 19.
- Zhang, Y., Jia, J., Ding, Y., Ma, Y., Shang, P., Liu, T. and Hu, G. (2016) Alpha-boswellic acid protects against ethanol-induced gastric injury in rats: involvement of nuclear factor erythroid-2-related factor hemeoxygenase-1 pathway.Journal of Pharmacy and Pharmacology. 2016;68 : 514–522.ISSN20427158, 00223573.doi: 10.1111/jphp.12532.
- Zhang, Y., Ning, Z., Lu, C., Zhao, S., Wang, J., Liu, B., Xu, X., Liu, Y.(2013). Triterpenoid resinous metabolites from the genus Boswellia: Pharmacological activities and potential species-identifying properties. Chem Cent J.7: 153. ISSN 1752153X .doi: 10.1186/1752-153X-7-153.
- Zhang, Z., Zhao, X. and Qin, X.(2017). Potential genotoxic and cytotoxicity of emamectin benzoate in human normal liver cells.Oncotarget. 8 : 82185– 82195.ISSN19492553.doi: 10.18632/oncotarget.18988.
- Zhou, S.F., Liu, J.P. and Chowbay, B. (2009).Polymorphism of human cytochrome P450 enzymes and its clinical impact. Drug Metab. Rev.41: 89–295. ISSN 10979883, 03602532.doi: 10.1080/ 036025309 02843483.
- Zhu, W.J., Li, M., Liu, C.,Qu, J.P., Min, Y.H., Xu, S.W. and Li, S.(2013). Avermectin induced liverinjury in pigeon: mechanisms of apoptosis and oxidative stress. Ecotoxicol. Environ. Saf. 98: 74-81. ISSN01476513, 10902414.DOI: 10.1016/j.ecoenv.2013.09.021.

النشاط المضاد للأكسدة لبعض المركبات الطبيعية في التخفيف من آثار السمية الكبدية التي يستحثها الإيمامكتين بنزوات في ذكور الفئران

شريفة فتح الله محمد داود1، طارق محمد مصطفي الأقرع² و أمينة محمد جاد زيدان¹ ¹قسم العلوم البيولوجية والبيئية- كلية الاقتصاد المنزلي- جامعة الازهر ²قسم الحيوان الزراعي والنيماتودا – كلية الزراعة – جامعة الازهر

الإيمامكتين بنزوات (EMB) هو مبيد حيوي يستخدم في الزراعة كمبيد حشري. ونظرًا لنطاق إستخدامة الواسع ، فمن السهل وصوله إلي البيئة وتأثيره على صحة الإنسان. لذلك تهدف هذه الدراسة إلى تقييم التأثير الوقائي للمركبات الطبيعية ضد السمية الكبدية التي يسببها EMB. هذه هي الدراسة الأولى التي تحدد التأثير الوقائي لهذه المستخلصات ضد تأثير ات EMB. تم عمل تحليلات كيمائية شملت قياس نشاط ADN ، CAT ، MDA ، انزيمات الكبدية التي يسببها SOD ، CAT ، MDA ، تم عمل تحليلات كيمائية شملت قياس نشاط ADN ، CAT ، MDA ، انزيمات الكبدية التعبير الجبني لحبيات EMB. تم عمل تحليلات كيمائية شملت قياس نشاط ADN ، CAT ، MDA ، انزيمات الكبد AST ، ALT ، ADN ، CAT ، MDA ، أملت دراسة قياس درجة SOD ، انزيمات الكبد *CAT ، Mos ، Caspase ، 11-1β ، caspases ، 21-1β ، Caspase ، 21-1β ، 212-1β ، 212-1*