

Journal of Agricultural Chemistry and Biotechnology

Journal homepage: www.jacb.mans.edu.eg
Available online at: www.jacb.journals.ekb.eg

Characterization of Soil-Indigenous Cyanobacterial Strains and Bioactivity Assessment

Randa M. Zaki¹; Ahlam A. M. Mehesen²; Eman H. Ashour¹ and Aida H. Afify^{1*}

¹ Microbiol. Dept., Fac. Agric., Mansoura Univ., Mansoura, Egypt.

² Agri. Microbiol. Dept, Agri. Res. Center (ARC) Sakha, Kafr ElSheikh, Egypt.



Cross Mark



ABSTRACT

Some cyanobacteria isolates were collected from soil samples at various locations from governorates of Kafr El-Sheikh and El-Dakahlia. By using morphological characterizations to identify these isolates. Heterocyst-forming cyanobacteria were *Anabaena* sp., *Nostoc* sp., *Oscillatoria* sp. and *Chroococcus* sp. Only, *Nostoc* sp. and *Anabaena* sp. isolates were represented at high frequency in the two isolated areas. The less frequency of occurrence in this study were *Oscillatoria* sp., *Chroococcus* sp., *Phormidium* sp., *Pseudoanabaena* sp. *Nostoc calicola* D and *Anabaena cylindrica* D were recorded as the most active cyanobacteria according to nitrogen fixation, nitrogenase activity, Indole Acetic Acid (IAA) production and dry weight determination. Therefore, it would be recommended to apply these strains of cyanobacteria in bio-organic farming.

Keywords: Cyanobacteria, nitrogen fixation, Nitrogenase activity, IAA

INTRODUCTION

The beneficial effects of cyanobacteria in paddy soils have recently received a lot of attention. The amount of nitrogen fixed by these blue-green algae may meet the nitrogen requirements of wheat plants. The release of growth-promoting substances during algal development should not be overlooked in this regard. Blue-green algae are a type of photosynthetic prokaryotes that can fix nitrogen from the atmosphere in submerged rice fields. The cyanobacterial process requires biological N₂ fixation, which is dependent on a sufficient population. The diversity of cyanobacteria ranges from unicellular to multicellular, branched filamentous to pigmented, autotrophic to heterotrophic, free-living to symbiotic, psychrophilic to thermophilic acidophilic to alkylotrophic, planktonic to epiphytic. Some workers have emphasized the importance of phytoplankton (Alam *et al.*, 1989).

Cyanobacterial strains have the ability to fix nitrogen in the fields of culture, food, and fertilizer (Muthukumar *et al.*, 2007). In Egypt, cyanobacterial strains were isolated, identified as *Anabaena* sp. and *Nostoc* sp. and used to provide rice crops with N₂ and made a high progress rice production (Afify *et al.*, 2018)

The present research aims to study the diversity, isolations and identification of cyanobacteria from different locations in Egypt and study their capability for production PGPR as IAA and nitrogenase enzyme as evidence for producing nitrogen available for plants and to increase the crop.

MATERIALS AND METHODS

Soil samples collection

For cyanobacterial isolation, air-dried soil samples collected from different sites in Kafr El-Sheikh and El-Dakahlia governorates were used. The properties of the soil samples were determined according to Piper (1950) and

Jackson (1973), as Mechanical analysis and obtained data are presented in Table (1).

Table 1. Physicochemical characteristics of the collected soil samples used in this study

Character	El-Dakahlia (Mansoura)	Kafr El-Sheikh (Sakha)
Physical analysis (%):		
Coarse Sand	2.24	1.55
Fine Sand	23.51	19.79
Silt	42.00	47.57
Clay	32.26	31.09
Soil Textural class	Clay loam	Clay loam
Chemical analysis:		
pH	7.97	7.9
EC (ds/m)	0.7	1.176
Cations (ppm)		
Ca ⁺⁺	68.01	122.13
Mg ⁺⁺	34.50	65.41
Na ⁺	397.17	646.85
K ⁺	8.33	25.07
Anions (ppm)		
CO ₃ ⁻	0	0
HCO ₃ ⁻	725.17	792.17
Cl ⁻	143.54	434.22
SO ₄ ⁻⁻	0	18.69

Isolation of cyanobacteria

To obtain the cyanobacterial isolates, sterilized 0.7% agarized of modified Watanabe medium were poured into Petri dishes (10 cm in diameter) according to El-Ayouty and Ayyad, 1972.

Purification of cyanobacteria

The unialgal cultures were purified as described by Pringsheim (1949).

Maintenance of cyanobacterial cultures

Stock cultures were maintained in a refrigerator at 5°C. on agar slants of Modified Watanabe Medium (El-Nawawy *et al.*, 1958).

* Corresponding author.

E-mail address: aidaafify@yahoo.com

DOI: 10.21608/jacb.2021.209209

Identification of the isolated cyanobacteria

Characterization and identification of the purified cyanobacteria isolates were carried out according to Castenholz (2015).

Selection of the most efficient N₂-fixing cyanobacteria strain

A growth curve experiment was conducted for 18 isolates to compare their growth activities and their capacities for N₂-fixation. Cyanobacteria isolates were cultivated for 30 days, to determine cyanobacteria dry weight, and the fixed nitrogen amount.

Preparation of standard cyanobacterial inoculum

The inoculum of the cyanobacterial identified strains was prepared by inoculating 500 ml Erlenmeyer flasks each containing 200 ml of Modified Watanabe liquid medium with a loopfull of 21 days old culture of each cyanobacteria strains. Inoculated flasks were incubated at 28-30°C under continuous illumination (2500 lux) for 21 days.

Determination of total nitrogen content

Total nitrogen in the cyanobacterial culture for each isolates were determined using the micro-Kjeldahl method according to Jackson (1973).

Nitrogenase activity

The capacity of cyanobacteria isolates to fix nitrogen was assayed by acetylene reduction technique according to Hardy *et al.* (1973). Nitrogenase activity was then calculated by the following formula

$$\mu\text{mole C}_2\text{H}_4/\text{ml culture} = \frac{R \times (\text{container volume (test tube)}/\text{incubation time} \times \text{Inj. vol.} \times D \times 22.4)}{10^3}$$

Where: Inj.= injecting volume – R = reading -D = The volume of the medium.

The results were presented as μ mole C₂H₄ / 100 ml culture /day.

Quantification of indole acetic acid (IAA) production

Each isolate was grown in its specific medium supplemented with 0.1% tryptophan according to Ahmad *et al.* (2005). Production of IAA in the supernatant was assayed using method described by Pilet and Chollet (1970). This method was shown to be most sensitive and most specific (Glickmann and Dessaux, 1995). The IAA concentrations were calculated from IAA standard curve (Salkowski Colorimetric Technique).

RESULTS AND DISCUSSION

Isolation, purification and identification of cyanobacteria

Out of the several isolates of cyanobacteria, from the rhizosphere soil of wheat plants grown in the different locations namely, kafr El-Sheikh, and El-Dakahlia governorates. Eighteen cyanobacterial isolates were obtained in pure cultures; bacteria free (El-Gamal *et al.*, 2008). These isolates were examined for their morphological and cultural characteristics (Table 2), according to Venkataraman (1981) & Roger and Ardales (1991), in liquid and solid Watanabe medium (Staub, 1961).

Table (2) also showed the distribution of these isolates in the two Governorates. Eighteen cyanobacterial isolates were identified up to genera. Results showed that *Anabaena* sp. were the dominating organisms.

Anabaena sp. were isolated at high frequency; where, six cyanobacterial isolates, three of them representing in kafr El-Sheikh Governorate and three representing in Dakahlia Governorate. Less numbers of cyanobacteria belonging to different genera namely *Phormidium* sp. (3), *Oscillatoria* sp.

(1), *Chroococcus* sp. (1) and *Nostoc* sp. (1) were also obtained in pure cultures from kafr El-Sheikh Governorate. But in El-Dakahlia Governorate *Nostoc* sp. and *Oscillatoria* sp. were isolated also at high frequency comparing to kafr El-Sheikh; where two isolates of each one of them were obtained. *Chroococcus* sp. isolates were equal where each governorate had one isolate. Zero numbers of cyanobacteria belonging to genus namely *Phormidium* sp. appeared in El-Dakahlia. On the other hand, appeared one isolate of *Pseudoanabaena* sp. which did not appear in kafr El-Sheikh.

Table 2. Occurrence of cyanobacterial genera in kafr El-Sheikh and El-Dakahlia soil samples.

Origen soil samples	Isolates Code	Cyanobacterial Genera	Isolates Frequency (%)
kafr El-Sheikh	N1		
	N3	<i>Anabaena</i> sp.	16.67
	K3		
	L3	<i>Nostoc</i> sp.	5.56
	H3	<i>Oscillatoria</i> sp.	5.56
	M1	<i>Chroococcus</i> sp.	5.56
	K5		
El-Dakahlia	O1	<i>Phormidium</i> sp.	16.67
	O3		
	B1		
	E2	<i>Anabaena</i> sp.	16.67
	E3		
	B2	<i>Nostoc</i> sp.	11.11
	F5		
	D2	<i>Oscillatoria</i> sp.	11.11
	D3		
	G3	<i>Chroococcus</i> sp.	5.56
	F2	<i>Pseudoanabaena</i> sp.	5.56
Total number of isolates		18	

Phenotypic properties of cyanobacterial genera and species

In the second edition of Bergey's Manual of Systematic Bacteriology (2001) cyanobacteria are subdivided into five quasi-taxonomic groups or subsections. Their diagnosis is based on the dichotomous key: the morphotype is unicellular or quasi-multicellular (trichome); division is binary or multiple (with formation of beocytes); presence or absence of differentiated cells (akinetes and heterocysts); and absence or presence of ramification (true or false) in the trichome (Pinevich, 2008 and Komárek *et al.*, 2014). All visible cultural characterizations and light microscopic preparations were observed and summarized in Table (3).

Determination some biological activities of cyanobacteria *in vitro*

1. Cyanobacteria' growth

Results illustrated in Fig (1) indicate that a great variation in biomass production between the different cyanobacterial genera and in some cases ecological effects are appeared. In El-Dakahlia governorate isolates, ranges of cyanobacterial mass production (g gL⁻¹ culture) during incubation periods (10, 20, 30 days) were found to be in the order *Nostoc* spp. (2 isolates) 0.4 to 3.0; *Anabaena* spp. (3 isolates) 0.2 to 3.0; *Oscillatoria* spp. (2 isolates) 0.3 to 2.1; *Chroococcus* sp. (1 isolate) 0.2 to 1.7, *Pseudoanabaena* sp. (1 isolate) 0.1 to 0.5 gL⁻¹ culture. However, in kafr El-Sheikh governorate isolates, ranges of cyanobacterial mass production (g/100 ml-culture) also were found to be in the order *Anabaena* (3 isolates) spp. 0.6 to 3.2, *Chroococcus* sp. (1 isolate) 1.1 to 3.5, *Phormidium* spp. (3 isolates) 0.1 to 2.5, *Nostoc* sp. (1 isolate) 0.4 to 2.0, *Oscillatoria* sp. (1 isolate)

0.1 to 0.5 gL⁻¹ culture. These results are in agreement with those obtained by (El-Zawawy, 2016 and Taha 2000) who found that cyanobacteria exhibited the highest dry weight of with increasing the incubation period.

Table 3. Summarized of cultural, morphological and microscopic characteristic of cyanobacteria isolated from soil revealed by microscopic observations

Isolates Code	Thallus color	Thallus morphology	Vegetative Cell			Site	Heterocysts			Akinetes			Cyanobacteria Identified Name
			Shape	Width (µm)	Length (µm)		Width (µm)	Length (µm)	Shape	Shape	Width (µm)	Length (µm)	
B2	Dark green	Gelatinous to rubbery	Cylindrical	2.3-2.7	2.7-5	Terminal and intercalary	3.2-4.1	3.8-5.9	Subspherical	Spherical or subspherical	3.6-5.4	2.7-6.3	<i>Nostoc calcicola</i>
N3	Green	Filaments	Barrel	4	5	Terminal, intercalary	4-5	4-5	Subspherical conical	3-6 in series, sub-spherical	5-6	6-7	<i>Anabaena oryzae</i>
E2 & E3	Green	Filaments	Short angular	4-4.5	2-3.5	Intercalary	5-6	6.5-7.5	Barrel	-	-	-	<i>Anabaena cylindrica</i>
F5 & L3	Blue-green, green or brown; black and crusty when dry	Filaments	Subglobose to barrel	2-3	3-4	Intercalary or terminal	5-7	5-6	Globose or ellipsoid	Spherical to elliptical	10	10	<i>Nostoc pruniforme</i>
K3 & N1	Oliveaceous green, blue green	Filaments	Cylindrical	3.6-4.1	4.1-5	Terminal or intercalary	4.2-5	4.2-5.4	Spherical	Ellipsoidal or oblong	5.0-6.3	5-8.1	<i>Anabaena variabilis</i>
B1	Blue green	Filaments	Sub-spherical	5.7-7.8	5.7-7.1	Intercalary	5.6-7.1	6.5-7.1	Subspherical or nearly quadratic	Spherical	12.8-14.2	12.8-14.2	<i>Anabaena qelatinnicola</i>
K5 & O1 & O3	Dark blue green	Filaments	-	1.6-1.8	2.6-3	-	-	-	-	-	-	-	<i>Phormidium foveolarum</i>
F2	Moving by gliding movement	Filaments	Cylindrical	1.5-1.8	3.9-4.6	-	-	-	-	-	-	-	<i>Pseudanabaena qaleata</i>
G3 & M1	Dark blue green	Slimy-gelatinous	Spherical	3-4	3-4	-	-	-	-	-	-	-	<i>Chroococcus minor</i>
D2 & D3 & H3	Dark blue green	Trichome solitary and straight	Solitary	4-8.2	4-7.1	-	-	-	-	-	-	-	<i>Oscillatoria brevis</i>

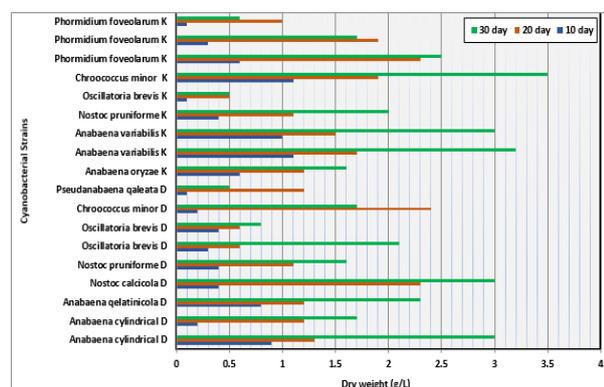


Fig. 1. Growth (g dry weight L⁻¹) of identified cyanobacterial strains isolated from El-Dakahlia (D) and kafr El-Sheikh (K) governorates

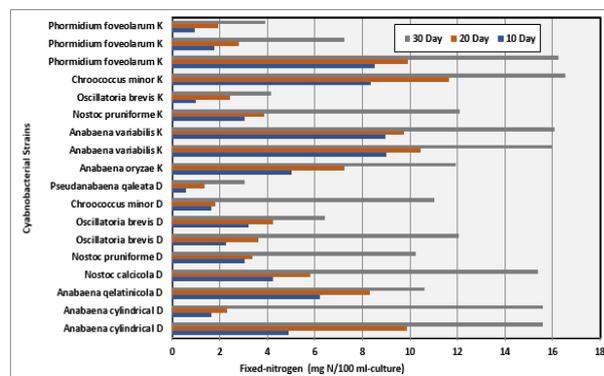


Fig. 2. Fixed nitrogen (mg N/100 ml culture) by cyanobacterial strains isolated from El-Dakahlia (D) and kafr El-Sheikh (K) governorates

2. Nitrogen fixation by cyanobacteria

It is important to determine the efficiency of cyanobacterial isolates to fix the atmospheric nitrogen as one of the most important parameters used for selection of cyanobacterial isolates in the further experiments.

Results graphically in Fig. (2) show that amounts of fixed nitrogen (mg N/100 ml culture) gradual increased by all tested cyanobacterial strains with incubation period increment, where higher values of the fixed nitrogen amounts were recorded with all strains at 30 days of growth. *Anabaena cylindrica* D and *Nostoc calcicola* D were recorded 15.60 and 15.37 mg N/100 ml liquid culture, respectively. On the other hand, only 3.06 mg N/100ml culture was recorded by the lowest one, *Pseudanabaena qaleata* D at 30 days incubation.

Firstly, in (1995) Stal stated that in nature cyanobacterial nitrogen fixation appears different patterns depending on the type of strain and environmental conditions. Boussiba *et al.*, (1984) found out that the use of cyanobacteria as a biofertilizer for rice fields is very promising but limited due to fluctuation in quality and quantity of inoculum and its physiological attributes in varied agroecological regions. Utilization efficiency of fixed nitrogen by rice plants is often low and efforts are therefore being extended to isolate suitable strains of cyanobacteria that would be prolific not only in fixing atmospheric nitrogen but also in excreting it continuously, thus making it available to the growing rice plants.

3. Cyanobacteria nitrogenase activity

The eighteen strains of cyanobacteria had various efficiency levels of the N₂-fixation as previously mentioned (Fig. 3). Thus, it is necessary to detect nitrogenase activity of these cyano-strains. Data illustrated in Fig (4) indicate that cyanobacterial strains had ability to reduced acetylene at varied rate from 20 to 540 n moles C₂H₄/ml/day. In addition, heterocystous cyanobacteria *Nostoc calcicola* and *Anabaena cylindrical* proved to be the most active strains for nitrogenase activity which recorded 540 and 360 n moles C₂H₄/ml/day, respectively. These findings in agreement with Singh et al., (2011) who reported that filamentous cyanobacteria have a specialized structure for nitrogen fixation called heterocysts which containing a key enzyme “nitrogenase” that is responsible and involved in the nitrogen fixation process.

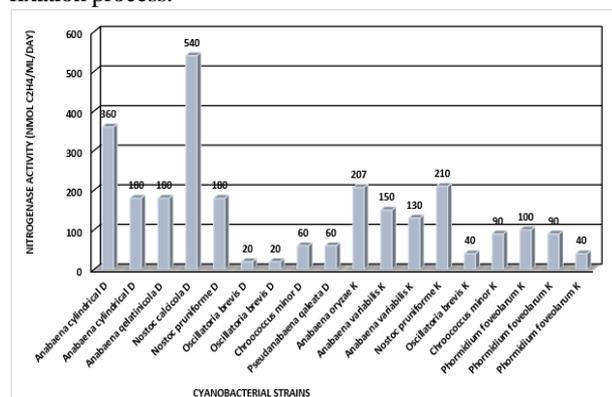


Fig. 3. Nitrogenase activity (n moles C₂H₄/ml/day) of cyanobacterial strains isolated from El-Dakahlia (D) and kafr El-Sheikh (K) governorates.

4. Production of indole acetic acid (IAA):

Results represent in Fig. (4) indicate that indole acetic acid production by tested cyanobacterial strains

through experiment age, 30 days. Most cyanobacterial strains showed positive results for IAA production in the media supplemented with tryptophan. It was also noted that through the first 10 days cyanobacterial strains had a high variable amounts then gradually decreased through next 10 days and some cyanobacterial strains became (Nil) IAA undetected at the end of incubation time (30 days). These results clearly demonstrated IAA production varied according to isolates that obtained from different environments. *Nostoc calcicola* D, *Anabaena cylindrica* D showed the superior significant production of IAA in first 10 days which were (59.85 mg /ml), (39.26 mg /ml) respectively.

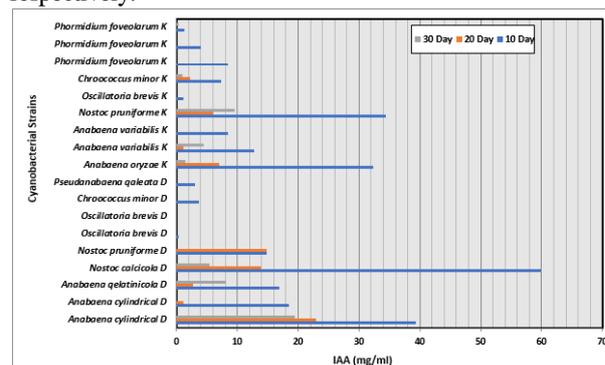
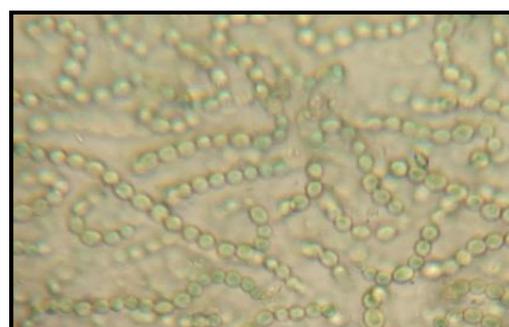


Fig. 4. IAA – production (mg /ml) of identified cyanobacterial strains isolated from El-Dakahlia (D) and kafr El-Sheikh (K) governorates.

It could be concluded that the cyanobacteria strains of *Anabaena cylindrical* D and *Nostoc calcicola* D (Fig. 5) were of the most efficient strains in biomass, IAA productions and nitrogen fixation capacity and could be used as bioagents for most economic plants.



Anabaena cylindrical



Nostoc calcicola

Fig. 5. Photographs of microscopic preparations of superior cyanobacterial strains.

REFERENCES

Afify Aida H.; F.I.A Hauka ; M.M. Gaballah and A.E.Abou Elatta (2018). Isolation and Identification of Dominant N₂ Fixing cyanobacterial strains from Different Locations. J. Agric.chem.and Biotechn., Mansoura Univ. 9(6):141-146.

Ahmad, F.; I. Ahmad and M.S. Khan (2005). Indole acetic acid production by the indigenous isolates of *Azotobacter* and fluorescent *Pseudomonas* in the presence and absence of treptophan. Turk. J.Biol.29:29-34.

Alam, M.J.; Habib, M. and Begum, M. (1989). Effect of water quality and dominance genera of phytoplankton on the abundance of available genera. Pak. J. Sci. Ind. Res. 32: 194-200.

Bergey's Manual of Systematic Bacteriology 2nd (2001). Rippka R., R.W. Castentholtz, I. Iteman and M. Herdman (2001). Form-genus I. *Anabaena* Bory. In: Boone, D.R. and W.R. Castenholz, (Eds.). Springer, Berlin, pp. 566–568.

- Boussiba S., Liu X.Q., Gibson J. (1984). Exogenous ammonia production by *Anacystis nidulans* R-2 induced by methionine sulfoximine. Arch. Microbiol., 138: 217-219.
- Castenholz, Richard, W. (2015). Bergey's Manual of Systematic Bacteriology "General characteristics of the Cyanobacteria. pp. 475.
- El-Ayouty, E.Y. and Ayyad, M.A. (1972). "Studies on blue-green algae of the Nile Delta 1-Description of some species in a wheat field". Egypt. J. Bot., 15:283-321.
- El-Gamal, A.D.; Ghanem, N.A.E.; El-Ayouty, E.Y. and Shehata, E.F. (2008). Studies on soil algal flora in Kafr El- Sheikh Governorate, Egypt. Egyptian J. Phycol. 9: 1-23.
- El-Nawawy, A. S.; Lotfi, M. and Fahmy, M. (1958). Studies on the ability of some blue-green algae to fix atmospheric nitrogen and their effect on growth and yield of paddy. Agric. Res. Rev. 36(2): 308-320.
- El-Zawawy, H.A.H. (2016). Microbiological and Ecological Studies on The Activity of Cyanobacteria in Different Types of Soil. PH.D Agric. Agric. Micro. Al- Azhar Univ.
- Glickmann, E. and Y. Dessaux, (1995). A critical examination of the specificity of the Salkowski reagent for indolic compounds produced by phytopathogenic bacteria Appl. Environ. Microbiol., 61(2):793-796
- Hardy, R.W.F.; R.C. Burs and R.D. Holsten (1973). Application of the acetylene-ethylene assay for measurement of nitrogen fixation. Soil Biol. Biochem., 5:47-81.
- Jackson, M.L. (1973). "Soil Chemical Analysis, Constable and CO₂". Agric. Exper. Mad. Wisconsin. USA.P: 183-187.
- Komárek, J., Kaštoký, J., Mareš, J. & Johansen, J.R. (2014). Taxonomic classification of cyanoprokaryotes (cyanobacterial genera) using a polyphasic approach. Preslia 86: 295-335.
- Muthukumar, C.; Muralitharan, G.; Vijayakumar R.; Panneerselvam, A. and Thajuddin, N. (2007). Cyanobacterial biodiversity from different freshwater ponds of Thanjavur. Tami Nadu (India). Acta Bat. Malacitana, 32:17-25.
- Pilet, P.E. and R. Chollet (1970). Sur le dosage colorimétrique de l'acide indolylacétique. C.R. Acad. Sci. Ser. d., 271:1675-1677.
- Pinevich, A. V. (2008). Paradoxes of Biodiversity, Phylogeny, and Taxonomy of cyanobacteria. Moscow University Biological Sciences Bulletin, 63 (1): 21-24.
- Piper, C. S. (1950). "Soil and plant analysis Inter. Science Publisher, Inc. New York, USA".
- Pringsheim, E.G. (1949). "Pure culture of algae, their preparation Maintenance". Cambridge. Univ.
- Roger, P.A. and Ardales, S. (1991). "Blue-Green algae collection". International Rice Research Institute, IRRI, Pub. Manila, Philippines.
- Singh, R.P., Baudh, K., Sainger, M., Singh, J.S., and Jaiwal, P.K. (2011). "Nitrogen use efficiency in higher plants under drought, high temperature, salinity and heavy metal contaminations," in Nitrogen Use Efficiency in Plants, eds V. Jain and A.P. Kumar (New Delhi: New India Publishing Agency), 99-123.
- Stal, L.J. (1995). Physiological ecology of cyanobacteria in microbial mats and other communities. New Phytol. 131, 1-32.
- Staub, R. (1961). "Ernährungsphysiologisch aurakologische untersuchungen an der planktischen blualg Oscillatoria rubescence D. C". Schweiz. Zeitschr Hydrobiologie, 23:82-198.
- Taha, M.T. (2000). "Biotechnological application of cyanobacteria in the constitution of a soil model of a biofortified farming system". M.Sc. Thesis, Fac. Sci., Azhar Univ., Egypt.
- Venkataraman, G.S. (1981). "The cultivation of algae". Indian Council Agric. Res., New Delhi, India.

توصيف سلالات السيانوبكتيريا المتوطنة في التربة وتقييم النشاط الحيوي

راندا محمد زكي السعداوى¹، أحلام علي مصطفى محيسن²، إيمان حسين عاشور¹ وعائدة حافظ عفيفي عامر^{1*}

¹ قسم الميكروبيولوجي - كلية الزراعة - جامعة المنصورة - مصر

² قسم الميكروبيولوجيا الزراعية - مركز البحوث الزراعيه - سخا - كفر الشيخ - مصر

تم جمع عينات تربيته من مواقع مختلفه في محافظتى كفر الشيخ والدقهليه وذلك لعزل السيانوبكتيريا حيث تم الحصول على عزلات السيانوبكتيريا نقيه ثم تعريفها وذلك طبقا شكل و لون الثالوس وحجم الهنتيروسست بالإضافة إلى الخلايا الخضريه و التكاثرية ، وعند تمييزها اتضح أن اجناس السيانوبكتيريا لها القدرة على تكوين الهنتيروسست مثل أنابينا و نوستوك و أوسيلاتوريا وكذلك كرووكوكس والفورميديم وسيدوانابينا. وكانت معظم الأنواع تتبع جنسى النوستوك و الأنابينا و ذلك في محافظتى كفر الشيخ والدقهليه بينما كانت أقل الأنواع تنتمى إلى جنسى الأسيلا توريا و الكرووكوكس اما الفورميديم يندعم وجودها في الدقهلية بينما السيدوانابينا لم تتواجد في أراضي كفر الشيخ وطبقا لتقديرات النيتروجين المثبت و الوزن الجاف و الاندول اسيتيك اسيد وانزيم النيتروجينيز. يتضح من النتائج المتحصل عليها ان النوستوك كالكولا والانابينا سيلانديكا هما أفضل العزلات التي يمكن أن يوصى باستخدامهما وذلك بالتطبيق في التجارب الزراعية لدعم الاتجاه الى الزراعة الحيوية والعضوية النظيفة.